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Assessing the health risk of living near composting facilities on lung health, fungal and bacterial disease in cystic fibrosis: a UK CF **Registry study**

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Abstract

Aim: To explore the health risk of living near permitted composting sites (PCSs) on disease severity in children and adults with cystic fibrosis (CF) across the UK.

Methods: A semi-individual cross-sectional study was used to examine the risk of disease severity in people with CF (pwCF) within and beyond 4 km of PCSs in the UK in 2016. All pwCF registered in the UK CF Registry were eligible for this study. Linear and Poisson regressions, adjusted for age, gender, genotype, BMI, Pseudomonas aeruginosa and deprivation, were used to quantify associations between distance to a PCS and percent predicted forced expiratory volume in one second (ppFEV₁), pulmonary exacerbations (#IVdays), and fungal and bacterial infections.

Results: The mean age of the 9,361 pwCF (3,931 children and 5,430 adults) studied was 20.1 (SD = 14.1) years; 53.3% were male; and 49.2% were homozygous F508del. Over 10% of pwCF (n = 1,015) lived within 4 km of a PCS. We found no statistically significant difference in ppFEV₁ and #IVdays/year in children. However, in adults, ppFEV₁ was -1.07% lower (95% confidence interval (CI): -2.29%, 0.16%) and #IVdays/year were 1.02 day higher (95%CI: 1.01, 1.04) within 4 km of a PCS. Furthermore, there were statistically significant differences in mean ppFEV₁ in CF adults with Aspergillus fumigatus (58.2.% vs 62.0%, p = 0.005) and Candida spp. (56.9% vs 59.9%, p = 0.029) residing within 4 km of a PCS. No associations were identified for allergic bronchopulmonary aspergillosis, P. aeruginosa or Staphylococcus aureus.

Conclusions: This novel national study provides evidence that adults with CF living near a PCS may experience small reductions in lung function, an increased risk of pulmonary exacerbations, and more frequent fungal infections. If confirmed by studies using refined exposure assessment methods accounting for bioaerosol dispersion, these results could have important implications for the living environment of pwCF.

Keywords: Cystic fibrosis, ppFEV₁, Pulmonary exacerbation, Fungal infection, Composting sites, Bioaerosol, ABPA

Background

Cystic fibrosis (CF) is an autosomal recessive disorder caused by mutations in the CF transmembrane conductance regulator (CFTR) gene which is located on the long arm of chromosome 7. It is one of the most commonly inherited life-shortening diseases in Caucasians with a birth prevalence of one in every 2,500 live births in the

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Respiratory infections are one of the leading causes of morbidity and mortality among the CF population. Substantial work has been done on the treatment and prevention of bacterial infection in pwCF [4, 5]. This has contributed to measurable increases in life expectancy but also may result in an increased risk for the colonisation of fungi [4]. Fungi are ubiquitous organisms and the most common fungus among pwCF is Aspergillus fumigatus [6]. Based on data from the UK CF Registry, the prevalence of positive cultures for A. fumigatus increased from 6% in 2007 to 15% in 2016 [1]. Evidence suggests that some fungal infections are associated with a greater decline in lung function and an increased rate of PEx [7]. The mechanisms underlying this trend remain unclear, but a substantial contribution from environmental factors seems likely [8, 9].

A better understanding of the risk of infections around composting facilities might help to better identify important environmental risk factors and their impact on the health of pwCF [10]. There are some public health concerns related to waste composting as it results in airborne bioaerosols which are released during the composting process [11]. The number of permitted composting sites (PCSs) more than doubled between 2010 and 2017 in England [12]. Concentrations of bioaerosols are higher at PCSs when the compost is agitated (e.g. shredded, turned and screened) [13]. These bioaerosols include fungi and fungal spores, gram-negative and spore-producing grampositive bacteria (e.g., actinomycetes), endotoxins and other various particulates sized between 0.2 to 100 µm in diameter [11]. The majority of bioaerosols emitted from composting facilities are small (<3 μ m in diameter) and therefore can be inhaled and penetrate the lungs [10]. Although previous studies have shown that bioaerosol concentrations fall within 250 m below levels considered acceptable by the UK Environment Agency's risk assessment [14], there is evidence that some bioaerosols may not ground over distances of a few kilometres from a PCS [15]. Furthermore, the role of these bioaerosols on the health of vulnerable population sub-groups remains unclear. It should be noted that, at present, there are no

tion on bioaerosol emissions from PCSs. Occupational studies have shown that exposure to fungi, bacteria and particulates, especially those $< 10 \ \mu m$ in diameter, affects human health [16]. These bioaerosols can penetrate the alveolar sacs of the lung, causing damage and loss of lung function, resulting in respiratory complications. These complications include allergic asthma, rhinitis, hypersensitivity pneumonitis, allergic bronchopulmonary aspergillosis (ABPA), eye and skin irritations, bronchitis, airway obstruction such as chronic obstructive pulmonary disease (COPD), organic dust toxic syndrome and toxic pneumonitis. The evidence of the possible associations between bioaerosol emissions from composting sites and health effects in the surrounding community is limited, although potential risks cannot be completely ruled out [10, 17, 18].

quantitative dose-response estimates to inform legisla-

People with lung diseases such as COPD, asthma and CF are likely to be more vulnerable to the harmful effects of living near a PCS, and exposure to bioaerosols may represent a particular health risk among pwCF [19]. This national-scale study aimed to investigate whether proximity to PCS—as a bioaerosol exposure proxy—is associated with a lower lung function, higher number of PEx, and more frequent fungal and bacterial infections in pwCF. This is based on the hypothesis that bioaerosols emitted from PCSs may be a source of such pathogens, including *A. fumigatus*.

Methods

Study design and population

We used a semi-individual cross-sectional study design to investigate the impact of living near a PCS on lung function, PEx, and fungal and bacterial disease among children and adults with CF. Our health outcome data came from the UK CF Registry; a secure database that collects annual data on approximately 10,000 patients from all CF specialist centres across the UK, representing>99% of pwCF [20]. Registry records are based on annual assessments of each individual, including data on demography, health outcomes and lifestyle factors, collected at CF centres across the UK, which are then centralised into one big database at the UK CF Registry. The present study focuses on all pwCF registered in the UK CF Registry who had an annual review conducted in 2016. In the UK, pwCF transition from paediatric to adult care occurs at around the age of 16. Individuals aged less than 16 years (0-15) were therefore considered children, and those aged 16 and above as adults. Most analyses were conducted separately for children and adults. UK CF Registry data accessible to the authors for this study were fully anonymised and stored and processed on a secure server with restricted and controlled access in compliance with Registry requirements and data confidentiality and privacy laws. Each individual was located based on their full residential postcode (postcode unit). Postcodes are regularly checked and updated in the UK CF Registry. In England and Wales, a postcode unit has an average of 43 (SD=38, median=33) residents and 18 (SD=15, median=14) occupied households [21]. Coordinates of postcode units were obtained from the Ordnance Survey Code-Point Open dataset [22].

Outcome variables

We considered three key outcome variables for this analysis: i) lung function, ii) PEx and iii) evidence of fungal and bacterial disease. Lung function was measured as the percent predicted forced expired volume in one second ($ppFEV_1$) as this is a key prognostic measure in CF [23]. $ppFEV_1$ was calculated using the Global Lung Function Initiative predictive equation [24]. Children under the age of six were included in the PEx analysis, but not in the lung function analysis (see below). Figure 1 provides a schematic overview of our inclusion and exclusion criteria.

There is no standard definition of a PEx, however, the use of intravenous (IV) antibiotics (which are prescribed to treat a PEx) is widely considered to be a good proxy in observational studies. Here, a PEx was defined as the use of IV antibiotics at home or in the hospital. We used the total number of days on IV antibiotics (home and hospital IV courses) received for a PEx during the twelve months preceding the annual review as we considered it to be a more robust and reliable measure of disease severity than the number of antibiotic courses [25]. Spirometry is not reliable under six years of age (i.e. < 6 years old) due to the inability of the child to perform the standard pulmonary function tests accurately [26]. Therefore, for our analysis of $pFEV_1$, all pwCF younger than six years



old (n=1,561) were excluded. Individuals who had a lung transplant before 2017 (n=296) were also excluded as their transplanted lungs and immunocompromised state would confound results.

To define the presence of fungal infection or disease, we used records of positive fungal cultures (sputum, cough/ throat swab or bronchoalveolar lavage) of *A. fumigatus* and *Candida spp*. in the year preceding the annual review and/or a record of ABPA. In the UK CF Registry, ABPA is defined using standard international diagnostic criteria (see Additional file 1: Appendix A) [27]. We also considered two common bacterial infections recorded in the UK CF Registry: *Pseudomonas aeruginosa* and *Staphylococcus aureus* using a positive record of culture in the year preceding the annual review.

Exposure variable

In the UK, commercial-scale composting facilities need a permit to operate. In England, such a permit is required if the facility is dealing with over 60 tonnes of compost at any one time [28], but different tonnages are used in Scotland, Wales and Northern Ireland. All operational PCSs in the UK at the end of 2016 were identified using the information provided by the UK Environment Agency (England), Natural Resources Wales, the Scottish Environment Protection Agency and the Northern Ireland Environment Agency. The sites were geocoded in a Geographic Information System (ArcGIS 10.7.1, ESRI, Redlands, CA) using the postcode of the facilities provided by the Environment Agencies. Figure 2 illustrates the distribution of PCSs across the UK. Distance between the nearest PCS and the pwCF's postcode was measured using Euclidian distances (point distance analysis) calculated in ArcGIS. In line with previous studies [10, 17, 18], we considered a range of circular distance bands $(0 - 250 \text{ m}, > 250 - \le 750 \text{ m}, > 750 - \le 1.5 \text{ km}, >$ $1.5 \le 2.5 \text{ km}, > 2.5 \le 4.0 \text{ km} \text{ and} > 4.0 \text{ km})$ around each PCS (Table S1 and Additional file 2: Appendix B). Ultimately, because the small number of pwCF living near PCS limited the statistical power of analyses in smaller bands, we focussed our study on within and beyond 4 km. The 4 km distance was chosen based on distance bands used in previous studies published in peer reviewed journals that used distance as a proxy for bioaerosol exposure around composting sites and associations of health outcomes [18, 29]. There are only a small number of community exposure studies examining bioaerosols emitted from composting facilities, and whilst these studies suggest that bioaerosols return to background levels within 2.5 km of a composting facility, these studies have many limitations, as discussed in two systematic reviews [10, 30].

Covariates

We chose a priori the following variables as potential confounders: age, sex, body mass index (BMI), CFTR genotype, P. aeruginosa, urban-rural classification and socio-economic status (SES). We categorised the age variable into two groups: children (<16 years) and adults $(\geq 16 \text{ years})$ [31, 32]. We used a binary variable for gender (M vs F) in our analysis. BMI is an important parameter to assess the nutritional status of CF individuals. We used absolute BMI in adults and BMI percentile, calculated based on age, sex, height and weight, in children [33]. Given inconsistencies in the literature around BMI categories (underweight, normal weight and overweight), we opted for the use of the continuous variable which reflects incremental gains (and losses) based on the absolute value [34]. Maintaining a BMI above the 50th percentile for children and an absolute $BMI > 19 \text{ kg/m}^2$ in adults is associated with better outcomes in CF [35]. The Office for National Statistics (ONS) postcode classification was used to assign an urban or rural designation to each pwCF residential postcode (see Additional file 3: Appendix C) [36]. We used the 2011 UK Townsend deprivation score quintiles at the output area level (highest resolution available) as a proxy of the SES of pwCF [36, 37]; the first quintile (Q1) refers to the least deprived areas of the UK whereas the fifth quintile (Q5) corresponds to the most deprived. Infections with P. aeruginosa was defined by a positive culture in the year preceding the annual review.

CFTR genotype

Two ways of classifying CFTR mutations have commonly been used in the past. First, six (or seven) classes are based on protein synthesis and function [38]. This classification system has significant limitations as there is considerable overlap between the classes and not all mutations have been studied sufficiently to be able to classify them. Second, a simplified classification using the common F508del mutation status as the main discriminator - i.e., homozygous F508del, heterozygous F508del and 'other'. This classification does not fully take into account the clinical phenotype as the 'other' category covers a wide spectrum of disease severity. To improve on these classifications and make them more relevant to contemporary practice, we devised a new eight-group classification (Table S2) based on the different combinations of the recognised terms developed in parallel with new CFTR-targeted therapies - so-called CFTR-modulators: residual function (RF), minimal function (MF) and gating function [39, 40]. We believe that an added benefit of this new classification is that it aligns better with potential treatment options for patients.



Statistical analysis

Spatial and statistical analyses were performed using ArcGIS 10.7.1 (ESRI, Redlands, CA) and *R* 4.1.1 [41], respectively. Individual characteristics including age, sex, *CFTR* genotype class and *P. aeruginosa* status, and area-level descriptors such as deprivation, rural–urban classification and country were compared among pwCF residing within and beyond 4 km from a PCS using twoway ANOVAs. Separate regression models were built for

children and adults. A multiple linear regression analysis was run to explore the association between ppFEV₁ and distance ($\leq 4 \text{ km } vs > 4 \text{ km}$) from a PCS. Poisson regression was employed to explore the association between the total #IVdays for PEx and distance from a PCS. The models were adjusted in a forward-stepwise approach with covariates added one by one using the Akaike information criterion (AIC) [42]. The best model was identified based on the lowest AIC (Table S3). A *p*-value of 0.05 was

considered statistically significant and 95% corresponding confidence intervals (CIs) and p-values are reported. Fungal and bacterial infection rates for *A. fumigatus, Candida spp.,* ABPA, *P. aeruginosa and S. aureus* were calculated among pwCF within and beyond 4 km of a PCS. An ANCOVA was used to calculate adjusted mean ppFEV₁, separately in children and adults, for each of these five measurements of fungal and bacterial disease.

Ethics and approval

This study used individual-level data pseudonymised by the UK CF Registry, which has UK National Health Service (NHS) research ethics approval (REC ref: 07/ Q0104/2) and consent from each person for whom data are collected, including use of pseudonymized data in approved research. The use of the data was approved by the UK CF Registry Research Committee (Data Request Reference: 302).

Results

A total of 9,695 pwCF had an annual review in 2016. Data for 9,631 individuals, including 3,931 children and 5,430 adults, were available for our PEx and fungal and bacterial disease analysis. After the removal of children < 6 years old, 2,370 children remained for our ppFEV1 analysis. Tables 1 and 2 compare the demographic characteristics of pwCF residing within and beyond 4 km from a PCS. The average age of pwCF was 20.1 (SD = 14.1) years; 49% (n=4,588) were aged 16–40 years; 53.3% were male and 19.2% lived in rural areas. There was an almost equal proportion of pwCF across all quintiles of deprivation. The geographical distribution of pwCF by country was as follows: England = 7,675 (82.0%), Wales = 494 (5.3%), Scotland = 796 (8.5%) and Northern Ireland = 396 (4.2%) (Fig. 2). In terms of CFTR genotypes, 49.2% were homozygous for F508del, 24.3% had at least one MF mutation, 11.4% had an RF mutation and 6.2% had a gating mutation (Table S2). In 2016, there were 256 active PCSs in the UK (209 in England; 24 in Wales; 21 in Scotland; and 2 in Northern Ireland) (Fig. 2); 1,015 pwCF (10.8%) lived within 4 km of a PCS.

Lung function

Overall, the mean ppFEV₁ was 85.0 within 4 km of a PCS *vs* 86.1 beyond 4 km (p=0.391) and 62.2 within 4 km *vs* 64.3 beyond 4 km (p=0.044) in children and adults, respectively. The age (p=0.021), sex (p<0.001), deprivation index (p=0.002), rural proportion (p=0.001) and *CFTR* genotype (p=0.001) characteristics of pwCF residing within 4 km from a PCS were all statistically significantly different to those living farther away from such facilities (Table 1). There was no statistically significant

difference in mean ppFEV₁ between children 6–15 years old with CF who lived within and beyond 4 km of a PCS (Table 3). In our unadjusted linear regression analysis, we found a -1.44% (95% CI: -2.84, -0.04) lower ppFEV₁ in adults with CF residing within 4 km of a PCS compared to the rest of pwCF. This difference in ppFEV₁ became smaller and the association was attenuated by -1.07% (95% CI: -2.29, -0.16, p=0.088) after adjusting for age, sex, BMI, genotype, *P. aeruginosa*, and deprivation (Table 3).

Pulmonary exacerbations

The mean number of #IVdays were 9.8 vs 9.5 days per year in children living within and beyond 4 km from a PCS whereas in adult the mean number of #IVdays were 22.4 vs 21.0 days per year. In univariate analyses, there was a statistically significant difference in #IVdays per year between males and females (p = < 0.003), deprivation quintiles (p=0.005), rural and urban areas (p=0.004), genotype category (p = 0.004) for pwCF residing within and beyond 4 km of the PCS (Table 2). We found no statistically significant difference in #IV days between children with CF who lived within 4 km of a PCS and those who lived farther away. Using Poisson regression analysis in adults, we identified an increase of 1.04 (95%CI: 1.03-1.06) and 1.02 (95%CI: 1.01,1.04) in the rate of #IVdays per year before and after adjustment for age, sex, BMI, genotype, P. aeruginosa and deprivation (Table 3).

Fungal and bacterial infections

Table 4 shows that there was no difference in the point prevalence of A. fumigatus positive cultures within and beyond 4 km of a PCS when considering all pwCF (15.3% vs 15.5%, respectively). However, there was a statistically significant difference in the mean $ppFEV_1$ in adults with CF with A. fumigatus positive culture within and beyond 4 km of PCS (55.9% vs 62.1%: p = 0.001). This difference remained significant after adjusting for age, sex, BMI, genotype category and deprivation (54.5% vs 59.7%; p = 0.001). Furthermore, the point prevalence of *Candida spp.* was 3.6% higher (p = 0.01) close to PCSs. The difference in mean ppFEV₁ increased to 4.2% (54.8% vs 59.0%; p = 0.042) in the adjusted multivariate analysis for adults. No statistically significant differences were found in the point prevalence of ABPA, P. aeruginosa or S. aureus within and beyond 4 km from a PCS in adults with CF. In addition, there was no statistically significant difference in the mean $ppFEV_1$ of children or adults with CF who live within and beyond 4 km of a PCS in the UK when considering cultures for ABPA, P. aeruginosa or S. aureus (Table 4).

Table 1	Mean ppFEV ₁	within and be	yond 4 km of a	a PCS in j	pwCF in the UK in	2016.	p-value < 0.	05 are shown	in bolc
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	Mean ppFEV ₁								
Characteristics	n (%)	mean (SD)	mean (<i>SD</i>)	<i>p</i> -value					
		<u>≤</u> 4 km	>4 km						
	n=7,800 (100%)	(<i>n</i> =866; 11.1%)	(n=6,934; 88.9%)						
Age (years)				0.0265					
Children < 6	_a								
6-15	2,370 (30.4)	85.0 (16.7)	86.1 (17.2)						
Adults \geq 16	5,430 (69.4)	62.2 (23.7)	64.3 (23.6)						
Sex				< 0.001					
Female	3,629 (46.5)	66.7 (24.5)	70.8 (23.8)						
Male	4,171 (53.5)	69.1 (24.0)	71.2 (24.3)						
Genotype				0.001					
Homozygous F508del	3,825 (49.0)	66.1 (24.5)	68.9 (24.0)						
F508del/MF	1,511 (19.4)	66.3 (23.2)	69.0 (24.1)						
RF/other	874 (11.2)	75.9 (24.6)	79.1 (22.5)						
F508del/other	558 (7.2)	70.3 (23.1)	73.7 (24.6)						
Gating/any ^b	494 (6.3)	73.3 (25.4)	75.9 (23.5)						
MF/MF	249 (3.2)	67.0 (18.5)	69.1 (21.9)						
Other/other	158 (2.0)	77.5 (21.9)	73.3 (23.6)						
MF/other	131 (1.7)	63.1 (28.3)	76.4 (22.8)						
P. aeruginosa culture				0.0016					
Negative	3,984 (51.1)	76.3 (22.3)	79.5 (21.7)						
Positive	3816 (48.9)	60.4 (23.5)	62.4 (23.3)						
Deprivation quintile (Townsend)				0.002					
Q1 (least deprived)	1,612 (20.7)	67.8 (23.6)	73.3 (23.6)						
Q2	1,615 (20.7)	65.8 (26.1)	71.1 (24.7)						
Q3	1,666 (21.4)	68.7 (22.9)	70.8 (24.1)						
Q4	1,547 (19.8)	69.2 (24.3)	70.3 (23.5)						
Q5 (most deprived)	1,360 (17.4)	68.3 (24.6)	69.3 (24.1)						
Rural and Urban				0.001					
Rural	1,500 (19.2)	66.2 (24.6)	72.4 (24.1)						
Urban	6,300 (80.8)	68.3 (24.2)	70.7 (24.0)						

ppFEV₁_percent predicted FEV₁, PCS permitted composting site, pwCF people with cystic fibrosis, Pseudomonas aeruginosa = P. aeruginosa

^a FEV₁ not reliable under 6 years of age hence excluded from ppFEV1 analysis; MF = Minimal function (a mutation that produces either no protein or protein that do not respond to currently approved *CFTR* modulators); RF = Residual function; Other = Either the mutation was not identified, or it was identified but classification into one of the other categories was not possible

^b Except RF mutation

Discussion

To our knowledge, this is the first national-scale study focusing on the impact of living close to a PCS on lung function, PEx, and fungal and bacterial disease in pwCF. There is limited prior evidence on the health impacts of living near PCSs in the general population or in sub-groups affected by respiratory diseases. Herr et al. found significant associations between bronchitis (OR = 3.59, 95%CI; 1.40, 9.47), waking up due to cough (OR = 6.59, 95%CI; 2.57,17.7) and coughing during the day (OR = 3.18, 95%CI; 1.24,8.36) among individuals living within 150-200 m of a PCS compared to individuals living farther away (>400-500 m) in Germany [43]. In Finland, Aatamila et al. also found more shortness of breath (OR 1.5, 95% CI 1.0–2.2) and tiredness (1.5, 1.1-2.0) among individuals living within distance zones of < 1.5 km from waste treatment centres (for municipal waste or composting) compared to individuals living within 3.0 to 5.0 km [44]. Various community and occupational studies have explored the impact of bioaerosols on respiratory health which have been summarised in a systematic review [30]. These studies have

Table 2 Mean #IVdays within and beyond 4 km o	a PCS in pwCF in the UK in 2016.	<i>p</i> -value < 0.05 are shown in bolc
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	#IVdays							
Characteristics	n (%)	mean (<i>SD</i>)	mean (SD)	<i>p</i> -value				
		\leq 4 km	>4 km					
	(<i>n</i> =9,361 (100%)	(<i>n</i> =1,015;10.8%)	(n=8,346; 89.2%)					
Age (years)				0.291				
Children < 6	1,561 (16.7)	5.5 (9.8)	5.1 (11.2)					
6-15	2,370 (25.3)	12.7 (21.8)	12.4 (20.9)					
Adults \geq 16	5,430 (58.0)	22.4 (34.2)	21.0 (32.7)					
Sex				0.003				
Female	4,371 (46.7)	22.0 (35.1)	18.7 (30.5)					
Male	4,990 (53.3)	14.2 (24.4)	13.9 (25.4)					
Genotype				0.004				
Homozygous F508del	4,603 (49.2)	22.9 (38.1)	17.7 (29.9)					
F508del/MF	1,822 (19.5)	12.3 (21.7)	12.4 (25.0)					
RF/other	1,070 (11.4)	8.4 (22.1)	8.4 (17.0)					
F508del/other	663 (7.1)	21.0 (30.1)	19.4 (30.1)					
Gating/any ^a	583 (6.2)	11.8 (19.6)	19.0 (27.6)					
MF/MF	298 (3.2)	18.7 (30.3)	11.0 (20.2)					
Other/other	176 (1.9)	10.6 (22.8)	12.2 (32.3)					
MF/other	146 (1.6)	5.7 (19.7)	6.2 (16.5)					
P. aeruginosa culture				< 0.001				
Negative	5,108 (55.2)	77.6 (22.4)	80.4 (21.4)					
Positive	4,139 (44.8)	61.2 (24.0)	62.8 (23.4)					
Deprivation quintile (Townsend)				0.005				
Q1 (least deprived)	1,905 (20.4)	15.2 (24.6)	14.1 (25.4)					
Q2	1,948 (20.8)	16.8 (27.1)	15.6 (28.1)					
Q3	1,982 (21.2)	15.7 (32.1)	15.4 (26.4)					
Q4	1,868 (20.0)	17.7 (30.5)	16.6 (28.8)					
Q5 (most deprived)	1,658 (17.7)	22.0 (33.6)	19.4 (31.5)					
Rural and Urban				0.004				
Rural	1,794 (19.2)	16.0 (25.1)	15.3 (26.4)					
Urban	7,567 (80.8)	18.1 (30.7)	16.3 (28.4)					

#IVdays Number of days on IV antibiotics for PEx, PCS permitted composting site, pwCF people with cystic fibrosis, Pseudomonas aeruginosa P. aeruginosa

MF = Minimal function (a mutation that produces either no protein or protein that do not respond to currently approved *CFTR* modulators); RF = Residual function; Other = Either the mutation was not identified, or it was identified but classification into one of the other categories was not possible

^a Except RF mutation

shown a possible increased airway irritation among residents living near composting sites. Other studies also found a significant association between organic dust and lung function [45, 46]. Nevertheless, Douglas et al. [17] did not find an increased risk of respiratoryrelated hospital admission among the general population living near PCSs in England, using distance from PCS as a proxy for bioaerosol exposure. In a follow-up study, Roca-Barcelo et al. [18] did not find an increased risk of being hospitalised with CF, nor any respiratoryrelated health outcome, in areas with higher exposure to *A. fumigatus* (estimated using dispersion models) near PCSs in England.

Although we found no associations for children with CF, our study found a relatively lower (1.07%) ppFEV₁ and higher (1.02 days/year) #IVdays among adults with CF living closer (≤ 4 km) to a PCS. To contextualise this, lower FEV₁ is strongly associated with higher mortality and poor quality of life, hence it is important to define the percentage of change in FEV₁ that is clinically relevant and significant in pwCF. FEV₁ has been used as the primary outcome in the majority of CF clinical trials due to its reproducibility and reliability [47, 48]. For example,

Table 3 Linear and Poisson regression analysis of the relationship between ppFEV₁ and #IVdays to residential postcode distance from PCS among children and adults with CF in the UK in 2016. *p*-value < 0.05 are shown in bold

Residential	ppFEV ₁					#IVdays					
postcode distance	Mean ppFEV ₁	Coeff	95% CI	p-value	AIC	mean #IVdays	Coeff	Exp (B)	95% CI	p-value	AIC
Children (6 -15 years) (<i>n</i> = 2,370)					Children (0 -15 years) (n = 3,931)						
Unadjusted				0.391	19,297.9					0.087	103,952.3
<u>≤</u> 4 km	85.0	-0.75	-2.46, 0.96			9.8	0.02	1.02	0.99, 1.05		
>4 km	86.1	1.00	-			9.5		1.00	-		
Adjusted ^a				0.358	18,887.6					0.623	84,834.0
<u>≤</u> 4 km	86.0 ^b	-0.74	-2.31, 0.83			10.3	0.01	1.01 ^b	0.99, 1.04		
>4 km	86.8 ^b	1.00	-			10.3		1.00 ^b			
Adults (16—86	years) (<i>n</i> = 5,430))				Adults (16—86 years) (n = 5,430)					
Unadjusted				0.044	47,438.5					< 0.001	231,519.0
<u>≤</u> 4 km	62.2	-1.44	-2.84, -0.04			22.4	0.04	1.04	1.03, 1.06		
>4 km	64.3	1.00	-			21.0		1.00	-		
Adjusted ^a				0.088	46,040.1					< 0.001	187,492.7
<u>≤</u> 4 km	64.4 ^b	-1.07	-2.29, 0.16			19.4	0.02	1.02 ^b	1.01, 1.04		
>4 km	66.3 ^b	1.00	-			18.4		1.00 ^b			

ppFEV, percent predicted FEV, #IVdays Number of days on IV antibiotics for PEx, pwCF people with cystic fibrosis, PCS permitted composting site

^a Adjusted for age, sex, BMI (children model adjusted for BMI percentile and adult model adjusted for absolute BMI), genotype, *P. aeruginosa* and deprivation ^b calculated using ANCOVA (Analysis of Covariance) adjusting for age, sex, BMI (children model adjusted for BMI percentile and adult model adjusted for absolute BMI), genotype, *P. aeruginosa* and deprivation

Wainwright et al. reported a significant improvement in ppFEV₁ ranging between 2.6–4.0% (p < 0.001) in phase 3, a randomized, double-blind study assessing the effects of lumacaftor/ivacaftor among 1,108 CF patients [47]. PEx are associated with a decline in FEV_1 and poor quality of life. Furthermore, they are considered a major cause of morbidity among pwCF. Although small reductions in mean $ppFEV_1$ and a one-day reduction in #IV days may be not clinically significant at an individual level, this could represent an important difference at the population level in the UK CF population. Environmental risk factors may play a critical role in the development of PEx with significant consequences and serious implications for the health of pwCF. A better understanding of these environmental risk factors, including how they mediate adverse effects and influence disease progression, could provide insight into the pathogenesis of respiratory disease in pwCF.

There was a significant reduction in mean $ppFEV_1$ (5.2%) and (4.2%) in CF adults with *A. fumigatus* and *Candida spp.* positive cultures living close to a PCS, respectively. Our study cannot tell us the origin of the pathogens, the potential mechanism underlying this, or indeed if it relates to an unmeasured factor. One possible explanation could be a higher sputum fungal burden (density) [15] close to PCSs. Williams et al. found high anthropogenic *A. fumigatus* modelled concentrations in a 3–4 km radius around some composting facilities in

England [15]. Although not statistically significant, living close to a PCS was associated with a higher prevalence of ABPA in children, which may provide some indirect evidence in support of this theory, as a higher density of fungal spores in the atmosphere could lead to greater sensitisation. This requires testing in robust prospective studies. Quantitative molecular microbiological methods to identify fungi at the species level would be able to elucidate fungal burden in sputum [49]. The majority of studies to date have only explored the association between lung function and positive sputum fungal cultures by traditional culture-based methods [50, 51]. For example, Amin et al. reported mean FEV₁ values of 79.2% and 86.1% among pwCF with Aspergillus positive and negative cultures, respectively (p = 0.04) [50], while Kraemer et al. found a decline in lung function in CF children with positive Aspergillus cultures compared to CF controls.

The major strengths of this UK-wide study are the large number of pwCF studied, the high quality of the UK CF Registry data, the availability of detailed individual-level residential information (full residential postcode) to assess distances, the inclusion of all registered PCS in the UK, the use of an improved genotype classification, and the inclusion of multiple covariates. This study aimed to consider proximity to PCS as a proxy for bioaerosol exposure and associations with selected outcomes in pwCF; therefore, we did not

	Distance ban	d		Distance band			
	Crude			Adjusted*			
	<u>≤</u> 4 km	>4 km	p-value	≤4 km	>4 km	p-value	
Total Number of pwCF (n)	866	6,934	-	-	-	-	
Number of pwCF who had culture** (n)	842	6,730	-	-	-	-	
A. fumigatus							
n (point prevalence %)	129 (15.3)	1,041 (15.5)	0.911				
mean ppFEV ₁							
Children	77.2	79.0	0.662	78.3	80.4	0.631	
Adults	55.9	62.1	0.001	58.2	62.0	0.005	
ABPA							
n (point prevalence %)	76 (8.8)	551 (7.9)	0.397				
mean ppFEV ₁							
Children	81.2	75.3	0.229	86.6	82.6	0.179	
Adults	60.2	59.1	0.717	60.3	60.1	0.696	
Candida spp.							
n (point prevalence %)	180 (21.4)	1,195 (17.8)	0.010				
mean ppFEV ₁							
Children	79.6	79.3	0.929	84.0	82.9	0.921	
Adults	53.6	57.8	0.048	56.9	59.9	0.029	
P. aeruginosa							
n (point prevalence %)	439 (52.1)	3,377 (50.2)	0.284				
mean ppFEV ₁							
Children	80.5	82.3	0.436	80.9	84.1	0.399	
Adults	56.8	58.1	0.280	58.5	59.6	0.240	
S. aureus							
n (point prevalence %)	76 (8.3)	558 (9.0)	0.469				
mean ppFEV ₁							
Children	71.2	76.1	0.541	63.7	70.8	0.472	

Table 4 Respiratory microbiology, fungal disease and mean ppFEV₁ among children and adults with CF within and beyond 4 km of PCS in the UK in 2016. p-value < 0.05 are shown in bold

58.0 Aspergillus fumigatus = A. fumigatus, Pseudomonas aeruginosa = P. aeruginosa, Staphylococcus aureus = S. aureus

* Adjusted for age, sex, BMI (children model adjusted for BMI percentile and adult model adjusted for absolute BMI), genotype and deprivation

** (Sputum/ Cough Swab/ Bronchoscopy); ppFEV1 percent predicted with FEV1, pwCF people with cystic fibrosis, ABPA Allergic Bronchopulmonary Aspergillosis, PCS permitted composting site; Children = 6-15 years; adult = 16-86 years

0.690

59.1

56.9

account for wind speed/direction, topography, or bioaerosol dispersion. We were also unable to account for background levels of bioaerosol (including seasonal variation), or other potential sources of anthropogenic bioaerosol (e.g., sewage treatments work, intensive farming). Future studies should consider using more detailed exposure assessment methods. There is also potential for further exposure misclassification as we geocoded PCSs based on postcodes provided by Environment Agencies which may not always reflect the exact location of the PCSs. We also did not differentiate between PCS by size or site type (e.g., whether the PCS is open windrow (outdoors) where emissions are

Adults

uncontained, or in-vessel (indoors) where emissions may be contained), which would likely influence exposure, as data on these characteristics provided by the Environment Agencies were not complete. Bioaerosols emitted from composting facilities are likely to return to background levels within a few kilometres [29]. Due to the small number of pwCF living within short distances, we were unable to investigate risk in this zone; nor within 250 m as no pwCF lived within this distance from a PCS. Therefore, we were unable to investigate risks where bioaerosol concentrations are likely to be the highest [17, 30, 52, 53]. Household measurements or close monitoring of the few pwCF living close to PCS

59.0

0.663

may provide quantitative evidence to support or adjust the UK Environment Agency's current risk assessment limit (250 m).

As with any registry study, there is the potential for bias, particularly by unmeasured factors. The small decline in ppFEV₁ the small increase in #IVdays and the associations found for fungal and bacterial disease in adults living within 4 km of a PCS may be due to residual confounding because the characteristics of those living near a PCS could be different from those living farther away. We limited our analyses to the most common fungal and bacterial species, so other potentially important clinical factors could have been missed. Further analyses should account, for example, for the use of antifungal treatment. We do understand that using chronic bacterial or fungal infections should be preferred over detecting them at least once a year. We also appreciate that inhaled antibiotics are a surrogate marker of chronic pseudomonas infection and Diabetes mellitus is considered a marker of severe disease. Due to the limited Registry data that were available for this project, we could not use chronic infection; and were unable to adjust our model for Diabetes mellitus and inhaled antibiotic therapy. Sputum results were based on UK CF Registry criteria and sampling frequency and technique (i.e., sputum, cough swab and bronchoscopy) will have varied (e.g., a higher proportion of children will have provided cough swabs). Further work including longitudinal sampling and molecular microbiological methods would be of benefit as this could help to: i) better characterise fungal infections experienced in pwCF; ii) investigate whether pwCF who have higher exposure to bioaerosols have an increased fungal burden density); and iii) explore the respiratory mycobiome in pwCF, including fungal diversity and its association with seasonal factors.

Conclusions

We found associations between proxies of disease severity in pwCF living within 4 km from a PCS (as a proxy for bioaerosol exposure) compared to those living farther away. The clinical relevance of these findings needs to be further assessed by studies with a better measure of bioaerosol exposure than the distance from the site, which accounts for bioaerosol dispersion, longitudinal sampling and molecular methods to identify and quantify pathogens in sputum. If our results are confirmed, the associations identified could have important implications on i) the living environments of pwCF, ii) clinical advice to pwCF; iii) public health advice provided to vulnerable populations living near PCS; and iv) how PCS are regulated and permitted.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12940-022-00932-1.

Additional file 1: Appendix A. Allergic Bronchopulmonary Aspergillosis and Candida spp.

Additional file 2: Appendix B. Distance bands.

Additional file 3: Appendix C. Rural-urban.

Additional file 4: Table S1. Number of people with CF (pwCF) living within specific distances from a permitted composting site (PCS) in the UK in 2016. Table S2. Classification of CFTR genotypes with frequencies among pwCF (n=9,361) in the UK in 2016. Table S3. Linear regression analysis of the relationship between ppFEV₁ and residential postcode distance from PCS among children and adults with CF in the UK in 2016. *p*-value <0.05 are shown in bold.

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Authors' contributions

FBP, NJS, and MSK conceived and designed the study. MSK performed the statistical analysis and drafted the original manuscript. All authors contributed to the interpretation of data discussed in the manuscript, revised the manuscript, and approved its final version to be published. FBP is the guarantor of this work and, as such, has full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis. The author(s) read and approved the final manuscript.

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Availability of data and materials

Health data can be obtained upon a formal request to the UK Cystic Fibrosis Registry (Apply for data from the UK CF Registry I Cystic Fibrosis Trust). The high resolution air pollution data can be obtained upon request from the Small Area Health Statistics Unit at Imperial College London (Small Area Health Statistics Unit | Faculty of Medicine | Imperial College London).

Declarations

Competing interests

NJS has received consultancy fees for advisory boards from Vertex, Chiesi, Gilead, Roche, Menarini and Pulmocide. He has also received speaker fees from Vertex, Chiesi, Gilead and Zambon. FBP has received consultancy fees from Vertex through Analysis Group Inc. The other authors have no conflict of interests to declare.

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References

- UK CF Registry. UK Cystic Fibrosis Registry Annual Data Report 2020. 2020. https://www.cysticfibrosis.org.uk/the-work-we-do/uk-cf-registry/reporting-and-resources. Accessed on 20th Feb 2022.
- 2. Clinical and Functional Translation of CFTR; 2022. https://cftr2.org/. Accessed 3 Mar 2022.
- Ranganathan SC, Skoric B, Ramsay KA, Carzino R, Gibson AM, Hart E, Harrison J, Bell SC, Kidd TJ. Australian Respiratory Early Surveillance Team for Cystic Fibrosis: Geographical differences in first acquisition of Pseudomonas aeruginosa in cystic fibrosis. Ann Am Thorac Soc. 2013;10(2):108–14.
- Gilligan PH. Infections in patients with cystic fibrosis: diagnostic microbiology update. Clin Lab Med. 2014;34(2):197–217.
- Chiappini E, Taccetti G, de Martino M. Bacterial lung infections in cystic fibrosis patients: an update. Pediatr Infect Dis J. 2014;33(6):653–4.
- Burgel PR, Paugam A, Hubert D, Martin C. Aspergillus fumigatus in the cystic fibrosis lung: pros and cons of azole therapy. Infect Drug Resist. 2016;9:229–38.
- Saunders RV, Modha DE, Claydon A, Gaillard EA. Chronic Aspergillus fumigatus colonization of the pediatric cystic fibrosis airway is common and may be associated with a more rapid decline in lung function. Med Mycol. 2016;54(5):537–43.
- Mendell MJ, Mirer AG, Cheung K, Tong M, Douwes J. Respiratory and allergic health effects of dampness, mold, and dampness-related agents: a review of the epidemiologic evidence. Environ Health Perspect. 2011;119(6):748–56.
- Crawford JA, Rosenbaum PF, Anagnost SE, Hunt A, Abraham JL. Indicators of airborne fungal concentrations in urban homes: understanding the conditions that affect indoor fungal exposures. Sci Total Environ. 2015;517:113–24.
- Robertson S, Douglas P, Jarvis D, Marczylo E. Bioaerosol exposure from composting facilities and health outcomes in workers and in the community: A systematic review update. Int J Hyg Environ Health. 2019;222(3):364–86.
- Douwes J, Thorne P, Pearce N, Heederik D. Bioaerosol health effects and exposure assessment: progress and prospects. Ann Occup Hyg. 2003;47(3):187–200.
- Douglas P, Fecht D, Jarvis D. Characterising populations living close to intensive farming and composting facilities in England. Front Environ Sci Eng. 2021;15(3):40.

- Taha MPM, Drew GH, Longhurst PJ, Smith R, Pollard SJT. Bioaerosol releases from compost facilities: Evaluating passive and active source terms at a green waste facility for improved risk assessments. Atmos Environ. 2006;40(6):1159–69.
- 14. Environment Agency and Department for Environment, Food & Rural Affairs. Risk assessments for your environmental permit; 2016. https://www.gov.uk/guidance/risk-assessments-for-your-environmental-permit. Accessed 3 Mar 2022.
- 15. Williams B, Douglas P, Roca Barcelo A, Hansell AL, Hayes E. Estimating Aspergillus fumigatus exposure from outdoor composting activities in England between 2005 and 14. Waste Manage. 2019;84:235–44.
- Mack SM, Madl AK, Pinkerton KE. Respiratory Health Effects of Exposure to Ambient Particulate Matter and Bioaerosols. Compr Physiol. 2019;10(1):1–20.
- Douglas P, Bakolis I, Fecht D, Pearson C, Leal Sanchez M, Kinnersley R, de Hoogh K, Hansell AL. Respiratory hospital admission risk near large composting facilities. Int J Hyg Environ Health. 2016;219(4–5):372–9.
- Roca-Barcelo A, Douglas P, Fecht D, Sterrantino AF, Williams B, Blangiardo M, Gulliver J, Hayes ET, Hansell AL. Risk of respiratory hospital admission associated with modelled concentrations of Aspergillus fumigatus from composting facilities in England. Environ Res. 2020;183:108949.
- Walser SM, Gerstner DG, Brenner B, Bünger J, Eikmann T, Janssen B, Kolb S, Kolk A, Nowak D, Raulf M, et al. Evaluation of exposure–response relationships for health effects of microbial bioaerosols – A systematic review. Int J Hyg Environ Health. 2015;218(7):577–89.
- Taylor-Robinson D, Archangelidi O, Carr SB, Cosgriff R, Gunn E, Keogh RH, MacDougall A, Newsome S, Schlüter DK, Stanojevic S, et al. Data Resource Profile: The UK Cystic Fibrosis Registry. Int J Epidemiol. 2018;47(1):9–10e.
- ONS. Office for National Statistics; Postcode Headcounts and Household Estimates - 2011 Census. 2013. https://www.nomisweb.co.uk/census/ 2011/postcode_headcounts_and_household_estimates. Accessed 22 Feb 2022.
- 22. Ordnance Survey. Ordnance Survey. 2022. https://www.ordnancesu rvey.co.uk/business-and-government/products/code-point-open.html. Accessed 08 March 2022.
- Szczesniak R, Heltshe SL, Stanojevic S, Mayer-Hamblett N. Use of FEV1 in cystic fibrosis epidemiologic studies and clinical trials: A statistical perspective for the clinical researcher. J Cyst Fibros. 2017;16(3):318–26.
- Quanjer PH, Stanojevic S, Cole TJ, Baur X, Hall GL, Culver BH, Enright PL, Hankinson JL, Ip MS, Zheng J, et al. Multi-ethnic reference values for spirometry for the 3–95-yr age range: the global lung function 2012 equations. Eur Respir J. 2012;40(6):1324–43.
- Hoo ZH, Campbell MJ, Curley R, Walters SJ, Wildman MJ. Do cystic fibrosis centres with the lowest FEV1 still use the least amount of intravenous antibiotics? A registry-based comparison of intravenous antibiotic use among adult CF centres in the UK. J Cyst Fibros. 2018;17(3):360–7.
- Kerem E, Viviani L, Zolin A, MacNeill S, Hatziagorou E, Ellemunter H, Drevinek P, Gulmans V, Krivec U, Olesen H. Factors associated with FEV₁ decline in cystic fibrosis: analysis of the ECFS Patient Registry. Eur Respir J. 2014;43(1):125.
- Stevens DA, Moss RB, Kurup VP, Knutsen AP, Greenberger P, Judson MA, Denning DW, Crameri R, Brody AS, Light M, et al. Allergic bronchopulmonary aspergillosis in cystic fibrosis–state of the art: Cystic Fibrosis Foundation Consensus Conference. Clin Infect Dis. 2003;37(Suppl 3):S225-264.
- Environment Agency. Waste exemption: T23 aerobic composting and associated prior treatment; 2014. https://www.gov.uk/guidance/wasteexemption-t23-aerobic-composting-and-associated-prior-treatment. Accessed 3 Mar 2022.
- 29. Williams B, Douglas P, Roca Barcelo A, Hansell AL, Hayes E. Estimating Aspergillus fumigatus exposure from outdoor composting activities in England between 2005 and 14. Waste Manag. 2019;84:235–44.
- Pearson C, Littlewood E, Douglas P, Robertson S, Gant TW, Hansell AL. Exposures and health outcomes in relation to bioaerosol emissions from composting facilities: a systematic review of occupational and community studies. J Toxicol Environ Health B Crit Rev. 2015;18(1):43–69.
- Vaz Fragoso CA, McAvay G, Van Ness PH, Metter EJ, Ferrucci L, Yaggi HK, Concato J, Gill TM. Aging-Related Considerations When Evaluating the Forced Expiratory Volume in 1 Second (FEV1) Over Time. J Gerontol A Biol Sci Med Sci. 2016;71(7):929–34.

- 32. Lalley PM. The aging respiratory system—Pulmonary structure, function and neural control. Respir Physiol Neurobiol. 2013;187(3):199–210.
- Vogel M. Vogel M. Childsds R package version 0.8.0 2022: data and methods around reference values in pediatrics; 2022. https://cran.r-proje ct.org/web/packages/childsds/index.html. Accessed 10 Mar 2022.
- Goss CH, Sykes J, Stanojevic S, Marshall B, Petren K, Ostrenga J, Fink A, Elbert A, Quon BS, Stephenson AL. Comparison of Nutrition and Lung Function Outcomes in Patients with Cystic Fibrosis Living in Canada and the United States. Am J Respir Crit Care Med. 2018;197(6):768–75.
- Stenbit AE, Flume PA. Pulmonary exacerbations in cystic fibrosis. 2011;17(6):442–7.
- Office for National Statistics (ONS). 2011 Census: aggregate data. [data collection]: UK Data Service; SN: 7427. https://doi.org/10.5257/census/ aggregate-2011-2. Accessed 3 Mar 2022.
- Taylor-Robinson DC, Smyth RL, Diggle PJ, Whitehead M. The effect of social deprivation on clinical outcomes and the use of treatments in the UK cystic fibrosis population: a longitudinal study. Lancet Respir Med. 2013;1(2):121–8.
- De Boeck K, Amaral MD. Progress in therapies for cystic fibrosis. Lancet Respir Med. 2016;4(8):662–74.
- Middleton PG, Mall MA, Dřevínek P, Lands LC, McKone EF, Polineni D, Ramsey BW, Taylor-Cousar JL, Tullis E, Vermeulen F, et al. Elexacaftor– Tezacaftor–Ivacaftor for Cystic Fibrosis with a Single Phe508del Allele. N Engl J Med. 2019;381(19):1809–19.
- Heijerman HGM, McKone EF, Downey DG, Van Braeckel E, Rowe SM, Tullis E, Mall MA, Welter JJ, Ramsey BW, McKee CM, et al. Efficacy and safety of the elexacaftor plus tezacaftor plus ivacaftor combination regimen in people with cystic fibrosis homozygous for the F508del</ em> mutation: a double-blind, randomised, phase 3 trial. The Lancet. 2019;394(10212):1940–8.
- R Core Team. R: A language and environment for statistical computing. Vienna: R Foundation for Statistical Computing; 2022. https://www.Rproject.org/.
- Burnham KP, Anderson DR. Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach. New York: Springer; 2003.
- Herr CEW, Zur Nieden A, Jankofsky M, Stilianakis NI, Boedeker RH, Eikmann TF. Effects of bioaerosol polluted outdoor air on airways of residents: a cross sectional study. Occup Environ Med. 2003;60(5):336–42.
- Aatamila M, Verkasalo PK, Korhonen MJ, Suominen AL, Hirvonen M-R, Viluksela MK, Nevalainen A. Odour annoyance and physical symptoms among residents living near waste treatment centres. Environ Res. 2011;111(1):164–70.
- Senthilselvan A, Chénard L, Grover V, Kirychuk SP, Hagel L, Ulmer K, Hurst TS, Dosman JA. Excess longitudinal decline in lung function in grain farmers. J Agromedicine. 2010;15(2):157–65.
- Jacobsen G, Schlünssen V, Schaumburg I, Taudorf E, Sigsgaard T. Longitudinal lung function decline and wood dust exposure in the furniture industry. Eur Respir J. 2008;31(2):334.
- Wainwright CE, Elborn JS, Ramsey BW, Marigowda G, Huang X, Cipolli M, Colombo C, Davies JC, De Boeck K, Flume PA, et al. Lumacaftor-Ivacaftor in Patients with Cystic Fibrosis Homozygous for Phe508del CFTR. N Engl J Med. 2015;373(3):220–31.
- Ramsey BW, Davies J, McElvaney NG, Tullis E, Bell SC, Dřevínek P, Griese M, McKone EF, Wainwright CE, Konstan MW, et al. A CFTR Potentiator in Patients with Cystic Fibrosis and the G551D Mutation. N Engl J Med. 2011;365(18):1663–72.
- Cuthbertson L, Felton I, James P, Cox MJ, Bilton D, Schelenz S, Loebinger MR, Cookson WOC, Simmonds NJ, Moffatt MF. The fungal airway microbiome in cystic fibrosis and non-cystic fibrosis bronchiectasis. J Cyst Fibros. 2021;20(2):295–302.
- Burgel P-R, Paugam A, Hubert D, Martin C. Aspergillus fumigatus in the cystic fibrosis lung: pros and cons of azole therapy. Infection and drug resistance. 2016;9:229–38.
- Kraemer R, Baldwin DN, Ammann RA, Frey U, Gallati S. Progression of pulmonary hyperinflation and trapped gas associated with genetic and environmental factors in children with cystic fibrosis. Respir Res. 2006;7(1):138.
- 52. Pankhurst LJ, Deacon LJ, Liu J, Drew GH, Hayes ET, Jackson S, Longhurst PJ, Longhurst JWS, Pollard SJT, Tyrrel SF. Spatial variations in airborne

microorganism and endotoxin concentrations at green waste composting facilities. Int J Hyg Environ Health. 2011;214(5):376–83.

 Deacon L, Pankhurst L, Liu J, Drew GH, Hayes ET, Jackson S, Longhurst J, Longhurst P, Pollard S, Tyrrel S. Endotoxin emissions from commercial composting activities. Environ Health. 2009;8(1):S9.

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